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Replace the paragraph beginning at page 14, line 27, with the following rewritten paragraph:

--The 1351-bp glutelin gene promoter region was PCR-amplified using rice genomic DNA as template and B1-5 (5'-GGGGAATTGATCTGATTTTGAGGAAT-3' (SEQ ID NO:3), EcoRI site underlined) as forward primer and B1-3 (5'-GGGGATCCCATAAGCTAT TTGTACTTGCT-3' (SEQ ID NO:4), BamHI site underlined) as reverse primer. The glutelin gene promoter plus 75-bp putative signal peptide sequence was PCR-amplified using rice genomic DNA as template and B1-5 as forward primer and B1-sp (5'-GGGGATCCGGGAT TAAATAGCTGGCCA-3' (SEQ ID NO:5), BamHI site underlined) as reverse primer. The truncated Apu encoding amino acid 106 to 1060 was PCR-amplified using genomic DNA of *T. ethanolicus* 39E as template and oligonucleotides 5'-CGGGATTCTTAAGCTTGCATCTTGA-3' (SEQ ID NO:6) (BamHI site underlined) as forward primer and 5'-CCGGCGGCCGCCTA CATATTTCCCCTGGCCA-3' (SEQ ID NO:7) (NotI site underlined) as reverse primer.--

Replace the paragraph beginning at page 15, line 9, with the following rewritten paragraph:

--The PCR-amplified GluB-1 promoter and GluB-1 promoter-signal peptide sequence were digested with EcoRI and BamHI and subcloned into the same sites in pBluescript (Strategene) to generate pBS-G and pBS-Gp. The truncated Apu was digested with BamHI and NotI and fused downstream of the GluB-1 promoter and GluB-1 promoter-signal peptide sequence in pBS-G and pBS-Gp, respectively, to make translational fusion and to generate pBS-G-Apu and pBS-Gp-Apu. The nopaline synthase gene germinator (Nos 3') was PCR-amplified using pBI221 (Clontech) as DNA template and oligonucleotide 5'-TCCGAGCTCCAGATCGT TCAAACATTT-3' (SEQ ID NO:8) (SacI site underlined) as forward primer and oligonucleotide 5'-AGCGAGCTCGATCGATCTAGAACAT-3' (SEQ ID NO:9) (SacI underlined) site as reverse primer. The Nos 3'UTR was digested with SacI and fused downstream of Apu in pBS-G-Apu and pBS-Gp-Apu to generate pBS-G-Apu-Nos and pBS-Gp-apu-Nos.--

Replace the paragraph beginning at page 15, line 20, with the following rewritten paragraph:

--The 1.2 kb promoter and signal peptide sequence of  $\alpha$ Amy8 was excised with SalI and HindIII from pAG8 (Chan et al., 1993, *supra*) and subcloned into pBluescript to generate PBS/8sp. The  $\alpha$ Amy8 3'UTRs was PCR-amplified using RAMYG6a as DNA template and oligonucleotide 5'-CGCCGCGGTAGCTTAGCTATAGCGAT-3' (SEQ ID NO:10) (SacII site underlined) as forward primer and oligonucleotide 5'-TCCCCGGGTCCTCTAAGTGAA CCGT-3' (SEQ ID NO:11) (SacII underlined) site as reverse primer. Plasmid RAMYG6a contains the 3' half portion of coding sequence and 3' flanking region of  $\alpha$ Amy8 genomic DNA and was generated by screening of a rice genomic DNA library (Clontech) using  $\alpha$ Amy8-C as a probe (Yu et al. (1992) Gene 122: 247-253). The  $\alpha$ Amy8 3'UTRs was subcloned into the SacII sites in PBS/8sp to generate PBS/8sp8U. The truncated apu was cut with BamHI and NotI and subcloned into the same sites in PBS-8sp8U to generate PBS- $\alpha$ Amy8-sp-Apu-8U.--

Replace the paragraph beginning at page 16, line 24, with the following rewritten paragraph:

--The truncated Apu encoding amino acids 106 to 1060 was PCR-amplified using genomic DNA of *T. ethanolicus* 39E as template and oligonucleotides 5'-CGCATATGTTAACG TTGCATCTTGATTTC -3' (SEQ ID NO:12) as forward primer and 5'-CCGCTCGAGCTAC ATATTTCCCCTGGCCA-3' (SEQ ID NO:13) as reverse primer. The amplified DNA fragment was digested with NdeI and XhoI and ligated into the same sites in pET20b(+) (Novagen) to generate pET-APU. pET-APU was transferred to *E. coli* strain BL21 (DE3) and APU was expressed. Purification of APU was performed according to the instruction provided by Novagen. One hundred micrograms of purified APU was injected into a New Zealand White rabbit successively at 4-6 week interval according to the methods described by Williams et al. (1995, Expression of foreign proteins in *E. coli* using plasmid vectors and purification of specific polyclonal antibodies, in: DNA Cloning 2-Expression Systems-A Practical approach. (Ed) Glover and Hames, IRL Press, New York).--